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<p>(54) Title: PRODUCTION OF HUMAN RECOMBINANT COLLAGEN IN THE MILK OF TRANSGENIC ANIMALS</p> <p>(57) Abstract</p> <p>Production of human procollagen or collagen in cells which ordinarily do not produce these molecules is effected by constructing expression systems compatible with mammary glands of non-human mammals. For example, expression systems can be microinjected into fertilized oocytes and reimplanted in foster mothers and carried to term in order to obtain transgenic non-human mammals capable of producing milk containing recombinant human procollagen or collagen. Human procollagen or collagen produced in this manner can be made of a single collagen type uncontaminated by other human or non-human collagens.</p>			

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PRODUCTION OF HUMAN RECOMBINANT COLLAGEN
IN THE MILK OF TRANSGENIC ANIMALS

This is a continuation-in-part of United States
Serial Number 08/011,643 filed 28 January 1993, the
5 contents of which are incorporated herein by reference.

Technical Field

The invention relates to production of recombinant
proteins, specifically collagen, in the milk of a
transgenic mammal. More specifically, it concerns
10 methods to prepare purified forms of useful human
collagen by effecting the secretion of the collagen (or
procollagen) into the milk of a transgenic mammal.

Background Art

Collagen is a major structural protein useful in
15 reconstructive therapeutic procedures in humans.
Collagens used for these purposes are generally prepared
by isolating the material from tissues of farm animals
such as cows or pigs. While such isolated collagen has
been used with some success, it is essentially a protein
20 foreign to the treated human being and immunogenic
responses can be a problem. This problem has been
minimized by treating the animal-derived collagen with
proteolytic enzymes to decrease immunogenicity.

It is clear that it would be advantageous to supply
25 human rather than bovine or porcine collagen for
therapeutic purposes. The sources for purified human
collagen are limited and the only reliable source is
human placenta. Human collagen can be purified from
human placenta as described in copending U.S. Patent
30 Application Serial No. 07/921,810 (Collagen Corporation).
The placenta contains several types of collagens, most
notably types I, III, IV, and V. The process of

separating and purifying one type from the others is imperfect and results in a predominant type with small amounts of the other types. Production of purified collagen from placentas further necessitates additional 5 processing steps to ensure that the resulting collagen product is free from human viruses such as hepatitis and HIV. In view of this, there have been attempts to prepare human collagen using recombinant techniques.

10 Expression of the human cartilage procollagen gene (Col2A1) in mouse 3T3 cells been reported (Ala-Kokko, L. et al., J Biol Chem (1991) 266:14175-14178). Olsen, A.S. et al. reported expression of a minigene version of the 15 human pro α 1(I) collagen gene in mouse fibroblasts (Olsen, A.S. et al., J Biol Chem (1991) 266:11117-1121). Full-length human pro α 2(V) collagen cDNA in pro α 2(V)-deficient hamster cells was reported by Greenspan, D.S. et al., J Biol Chem (1989) 264:20683-20687; mouse fibroblasts have also been used to express the pro α 1(I) chain wherein the resulting expressed protein is complexed in the collagen 20 triple helix with murine pro α 2(I) chains, as described by Schnieke et al., Proc Natl Acad Sci USA (1987) 84:8869-8873. Transgenic mice that were modified to contain a mutated form of the pro α 1(I) gene were not viable after birth, according to a study by Stacey, A. et al. Nature 25 (1988) 322:131-136. In addition, transgenic mice have been obtained that express a minigene version of the human gene for type I procollagen systemically (Khillan, J.S. et al., J Biol Chem (1991) 266:23373-23379); PCT application WO92/22333. These mice are useful as model 30 systems for investigating bone diseases characterized by the modified collagen produced.

35 The production of recombinant human collagen is made troublesome by the necessity for a multiplicity of posttranslational enzymes which are generally believed to be present only in cells which natively produce collagen.

At least eight such posttranslational enzymes are believed to be needed (Prockop et al., New England J Med (1984) 311:376-386). This has limited attempts at recombinant production to cells which natively produce 5 this protein; this inevitably results in chimeric forms of the protein.

In order to avoid chimeric collagens which contain partly human and partly host mammalian cell chains in the triple helix, it might be possible to use human cells for 10 this production. Even in this case, however, it is not possible to obtain collagen product of a particular type free of other collagen types. As further described below, the variety of collagens produced and their innate 15 similarity makes homogeneous preparations from either native or recombinant sources which produce their own collagen impossible.

The present invention solves these problems by effecting the synthesis of human procollagen or collagen in cells which do not natively produce this protein, 20 employing techniques established for the production of foreign proteins in mammalian milk, as described in the publications cited hereinbelow. The collagen of designated types is secreted into the milk either as 25 procollagen or collagen, depending on the construction of the expression systems and accompanying recombinant enzyme production.

Disclosure of the Invention

The invention provides recombinant production of 30 human collagen in a form that permits isolation of a homogeneous collagen type and can be designed to effect the production of commercially practical amounts of these proteins at a reasonable cost. The invention utilizes systems developed for the production of recombinant proteins in mammalian milk and requires utilization of

these techniques not only to effect the expression of the gene encoding the desired collagen, but also, if required, expression of the gene for any required posttranslational enzymes.

5 Thus, in one aspect, the invention is directed to a method for the recombinant production of human procollagen or collagen comprising recovering milk from the mammary glands of a nonhuman mammal. The mammal will have been modified to contain an expression system that 10 comprises DNA encoding human procollagen under the control of regulatory sequences operable in mammary glands. The human procollagen or collagen produced is recovered from the milk by various purification techniques. The nonhuman mammal may also be modified if 15 necessary to contain an expression system for the production of any needed posttranslational enzymes in the milk protein-secreting cells of the mammary glands.

Either collagen or procollagen may be secreted depending on the presence or absence of suitable 20 proteases in the cell. The procollagen encoded in the nucleotide sequence contained in the expression system will be preceded by a nucleotide sequence encoding an appropriate signal, either that natively associated with the procollagen or an alternate signal sequence workable 25 in the targeted cells. Thus, the procollagen produced as a result of a recombinant expression will be secreted into the milk. If the host cells contain enzymes which ordinarily effect cleavage of the prosequences from collagen -- i.e. procollagen N-protease and/or 30 procollagen C-protease, the procollagen will be cleaved of the prosequences as it exits the cell and collagen will be secreted into the medium. However, if these enzymes are absent from the production cell, procollagen itself will be secreted. Low levels of these proteases 35 will result in mixtures of collagen and procollagen.

Apparently the levels of these enzymes vary in cells which natively produce collagen. Depending on the tissue and the developmental stage of the subject from which the tissue originates, a greater or lesser proportion of 5 procollagen or collagen will be contained in the secreted materials. Thus, the milk which contains the collagen of the invention will contain this collagen in the form of human collagen *per se*, human procollagen *per se* or a mixture of both.

10 While it would be possible to modify the native procollagen genes to delete the coding sequences for the prosequences, it is not desirable to do this since the pro-region, especially the C-terminal pro-region, mediates the formation of triple helices by the collagen 15 portion of the molecule. Thus, if the prosequences are deleted from the expression vector, the resulting single collagen chains would be unable to form the triple helix which characterizes the collagen molecules.

20 If procollagen is secreted into the milk, of course, by supplying the appropriate proteolytic enzymes, collagen will result.

25 In another aspect, the invention is directed to expression systems useful in the foregoing method which comprise a DNA sequence encoding human procollagen operably linked to a promoter and other regulatory sequences capable of effecting expression in mammary glands. If necessary, expression systems operable in mammary glands for production of posttranslational enzymes can also be used. The invention also is directed 30 to nonhuman embryonic stem (ES) cells and to nonhuman eggs, including fertilized forms, modified to contain the expression system as well as to the nonhuman mammal implanted with the fertilized egg or with a blastula including the ES cells.

In other aspects, the invention is directed to milk containing human procollagen or collagen, and to homogeneous forms of human procollagen or collagen. These forms are made available by the practice of the 5 invention method which permits the production of only the recombinant collagen type desired absent a background of either similar nonhuman collagen molecules, or of collagens of different types.

Modes of Carrying Out the Invention

10 Collagen is a well studied protein, and the expression of genes encoding collagen has also been reviewed recently (Adams, S.L., Amer J Respir Cell and Molec Biol (1989) 1:161-168). This review summarizes the types of collagen known to occur and describes their 15 common features. The mRNAs encoding collagens of various types are translated in the cytoplasm of collagen-producing cells into procollagen subunits which are then assembled into triple helices. The assembled procollagen contains propeptide extensions at the N and C termini 20 that help to assemble the subunits, but do not participate in the triple helix. The prosequences are then cleaved to obtain collagen triple helix as the procollagen is secreted. The collagen helix itself contains nonhelical extensions designated telopeptides. 25 The triple helical regions contain repeating amino acid sequences with a glycine in every third position and proline (P) or hydroxyproline (HP) often in the other positions so as to contain a sequence of "triplets" of the form -(GXY)_n-, wherein X or Y or both are P or HP. 30 One of the essential posttranslational steps is the conversion of some proline residues to hydroxyproline to ensure stability of the triple helix at body temperature. Other important posttranslational modifications are disulfide exchange, hydroxylation of lysyl residues,

addition of carbohydrate and the assembly and crosslinking of the triple helical collagen molecules.

According to the Adams review, thirteen genetically distinct collagen types have been described and represent the products of at least 23 genes. The most common types found in interstitial tissues are types I, III, V and VI; in cartilage, types II, IX, X and XI are found. Some of these types exist natively as homotriplexes; others are heterotriplexes.

The nomenclature for the various collagen types is designed to designate the genetic origin of the collagen in question. For example, the triple helix of type I collagen is a heterotriplex containing the products of two different collagen-encoding genes. This type of collagen is designated $[\alpha_1(I)]_2, \alpha_2(I)$; thus, type I collagen triplets contain two chains encoded by the Col1A1 gene and one protein chain encoded by the Col1A2 gene. Type III collagen is designated $[\alpha_1(III)]_3$, and is thus comprised of three identical chains translated from the Col3A1 gene. Type II collagen is also a homopolymer designated $[\alpha_1(II)]_3$, which is comprised of translation products of the Col2A1 gene.

Since collagen-producing cells, as described above, produce several types of collagen, it has, in the past, been impossible to obtain, for example, homogeneous type I collagen free of type III collagen. By producing collagen in noncollagen-producing cells according to the method of the invention, obtaining such homogeneous preparations becomes possible.

The genetic materials for use in the method of the invention encoding the desired collagens are available. The genes encoding human types I, II, III, IV and V collagen are currently available.

Prockop, DJ et al. (supra) list the following cotranslational and posttranslational modifications that

occur when collagen is produced in fibroblasts: cleavage of signal peptides at the N-termini of the chains, hydroxylation of the Y-position proline and lysine residues, hydroxylation of a few X-position proline 5 residues, addition of galactose or galactose and then glucose to some of the hydroxylysines, addition of a mannose-rich oligosaccharide to the C propeptides, association of the C-terminal propeptides through a process directed by a structure of these domains, and 10 finally formation of both intrachain and interchain disulfide bonds in the propeptides. After secretion of the procollagen, the N propeptides are cleaved by a procollagen N proteinase and the C propeptides by a separate procollagen C proteinase. The collagen then 15 self-assembles into fibrils, and lysyl oxidase converts some lysine and hydroxylysine residues to the aldehyde derivatives to form cross-links with similar residues in adjacent molecules.

It is not entirely clear whether mammary cells, 20 since they do not endogenously produce collagen, contain the enzymes necessary for these posttranslational events. Since the assembly into triplexes is mediated by the sequences of the C-terminal extensions, in the event the epithelial cells of the mammary glands lack the required 25 proteases, it is believed that the assembly into triplexes can be effected extracellularly by providing appropriate secretion signals to the procollagen molecule as stated above and adding suitable proteases. Alternatively, the proteases could be produced 30 recombinantly in the epithelial mammary cells. The enzymes most likely to be needed by the mammary cells in order to effect required posttranslational processing are protein disulfide isomerase and the α -subunit of prolyl hydroxylase. If these enzymes are not endogenously 35 produced and must be provided recombinantly, expression

systems for their production may be supplied along with the expression systems for the collagen or procollagen itself. The gene for the α -subunit of prolyl hydroxylase has not yet been completely described but the gene 5 encoding the protein disulfide isomerase has been partially sequenced as described by Tasanen, K., et al, J Biol Chem (1988) 263:16218-16224 and J Biol Chem (1992) 267: 11513-11519. Genes encoding both proteins can be obtained using standard techniques. These two enzymes 10 function together as a tetrameric protein comprising two subunits of prolyl hydroxylase noncovalently associated with two α subunits of protein disulfide isomerase. Although the two subunits of protein disulfide isomerase are functional as a dimer, the two α subunits of prolyl 15 hydroxylase must be associated with protein disulfide isomerase in order to be active (Vuari, et al., 1992).

A well developed system for use in the invention method utilizes milk production in cows. This system is summarized by Krimpenfort, P. et al. in Biotechnology 20 (1991) 9:844-847. This article describes microinjection of fertilized bovine oocytes with genes encoding human proteins and development of the resulting embryos in surrogate mothers. The human genes were fused to the bovine α S₁ casein regulatory elements. This general 25 technology was also described in PCT Application WO91/08216 published 13 June 1991 and assigned to GenPharm.

Additional descriptions of the production of recombinant proteins by developing transgenic animals 30 which secrete the proteins into milk are found in European Application 264166 published 20 April 1988, assigned to Integrated Genetics. This disclosure emphasizes use of whey acid protein control systems to effect protein secretion and cites use of this system for the production of tPA and Hepatitis B surface antigen in 35

goat milk. Analogous systems for production of foreign proteins are described in PCT application WO88/00239 published 14 January 1988 and assigned to Pharmaceutical Proteins Limited. This application describes procedures for obtaining suitable regulatory DNA sequences for the products of the mammary glands of sheep, including beta lactoglobulin, and describes the construction of transgenic sheep modified so as to secrete foreign proteins in milk. An additional application, PCT 10 WO88/01648, published 10 March 1988 and assigned to Immunex Corporation, generally describes construction of transgenic animals which secrete foreign proteins into milk under control of the regulatory sequences of bovine alpha lactalbumin gene. Finally, PCT application 15 WO88/10118, published 29 December 1988 and assigned to Biogen, describes construction of transgenic mice and larger mammals for the production of various recombinant human proteins in milk. Other publications which describe the production of various proteins in milk 20 include Archibald, A.L. et al. Proc Natl Acad Sci USA (1990) 87:5178-5182 which describes production of human α -antitrypsin in the milk of transgenic mice. This production utilized a hybrid gene constructed from the β -lactoglobulin gene fused to an α 1-antitrypsin minigene. 25 Pittius, C.W. et al. Proc Natl Acad Sci USA (1988) 85:5874-5878 describe production of tissue plasminogen activator in the mammary glands of transgenic mice using the murine whey acidic protein promoter. Hennighausen, L, Protein Expression and Purification (1990) 1:3-8 30 provides a review of the use of the mammary gland as a bioreactor and the production of various foreign proteins in milk. This article describes the factors that affect the level of production and indicates recommended forms of expression system construction. The disclosures of

the foregoing publications are incorporated herein by reference.

Thus, techniques for construction of appropriate host vectors containing regulatory sequences effective to produce foreign proteins in mammary glands and cause the secretion of said protein into milk are known in the art. In addition, techniques for constructing transgenic mammals containing these systems, including mice as well as larger mammalian species such as cows, sheep and goats, are well known.

Systems for the expression of the procollagen gene in cells that produce milk protein can be constructed using methodology analogous to that recently described for the production of human collagenase in the lungs of transgenic mice (D'Armiento et al., Cell (1992) 71:955-961).

Genes encoding a number of procollagen types have been obtained; and genes for additional types can be obtained similarly. The preparation and cloning of the human Col1A1 gene has been described (Barsh et al., J Biol Chem (1984) 259:14906-14913). Briefly, a human genome cosmid library is packaged and used to transduce E. coli, which are plated, grown, and screened using a nucleic acid sequence specific for the Col1A1 gene. Positive colonies are located, matured in broth, and the DNA isolated. Restriction endonucleases are used to cut the DNA at selected sites. The digested DNA is examined by gel electrophoresis and DNA sequencing. A cosmid clone CG 103 isolated from a human genomic library was shown to contain the entire human Col1A1 gene.

Fragments of collagen genes have been selected from cosmid libraries (Barsh et al., supra) and from bacteriophage libraries (Chu et al., J Biol Chem (1985) 260:4357-4363 for type III collagen; Chu et al., Nature (1984) 310:337-340 for type I collagen). The Col1A1 gene

was obtained in three overlapping genomic clones using the Charon 4A bacteriophage vector. The Col1A2 gene has also been obtained from five overlapping clones in Charon 4A libraries (deWet et al., J Biol Chem (1987) 5 262:16032-16036). It has been shown that the first intron is important in regulating the $\alpha 1(1)$ gene expression in a tissue-specific manner in transgenic mice (Slack, J.L. et al. Mol Cell Biol (1991) 11:2066-2074).

As an alternative to using the entire gene, full-length cDNAs could be used, although the use of the entire gene has been shown to be more effective in transgenic animal experiments (Palmeter et al., Proc Natl Acad Sci USA (1991) 88:478-482). Such a full-length cDNA can be isolated from cDNA libraries, as was done for the cDNA for the alpha-2 chain of type I collagen (Lee et al., J Biol Chem (1988) 263:13414-13418), which was isolated from a lambda phage library.

To construct an expression system compatible with the epithelial cells of mammary glands, the Col1A1 or other procollagen gene, as a DNA fragment is ligated to a similarly prepared DNA fragment containing the promoter and any additional required regulatory sequences for a milk-specific protein expression. As described by D'Armiento et al., when ligating a promoter to a gene, it is necessary to preserve the translational start site for the protein. This may be accomplished by introducing a specific restriction endonuclease site immediately preceding the translation start site that is also unique for the 5' end of the chosen promoter. When these fragments are prepared using such a restriction endonuclease, the sites at the 3' end of the promoter will be compatible with the 5' end of the Col1A1 gene. When ligation occurs, the promoter will be ligated at the correct site of the gene to encode a messenger RNA that will allow translation from the translation start site of

the procollagen gene, analogous to ligation of the heptoglobin promoter to the human collagenase gene described by D'Armiento et al., supra. The promoter-gene construct is ligated into a bacteriophage vector cloning system by treating the phage DNA with a restriction endonuclease; both ends of the foreign DNA are then ligated to the vector construct for cloning the DNA.

CDNA containing the translation start site for expressed messenger RNA can also be ligated to a promoter to prepare a functional construct for introduction into a transgenic animal. This method was used for the human lactoferrin cDNA fused to the bovine alpha S1-casein gene 5' and 3' untranslated regions (Krimpenfort).

It is also understood that upstream regions of the promoter may be involved in regulating gene expression. Specifically, it has been shown that the extracellular matrix and hormones regulate the expression of bovine β -casein by their influence on the upstream sequences in the relevant gene (Schmidhauser, C. et al. Proc Natl Acad Sci USA (1990) 87:9118-9122). In addition, signals for termination of transcription and translation are also helpful in elevating levels of expression.

In order to reduce the size of the procollagen gene so that the construct could be cloned in bacteriophage, the gene itself could be shortened by reducing the size of the introns. This could be done for procollagen genes that are cloned as overlapping fragments. The introns at the junction sites of the fragments could be identified and treated with specific endonuclease to shorten the introns, but leave restriction sites that are compatible for ligation. Restriction sites could be altered by site-directed mutagenesis (D'Armiento et al., supra) to generate restriction sites for ligation of the fragments of the procollagen gene into a single construct. Another method of accomplishing the removal of introns is to

prepare fusion genes containing cDNAs to replace two or more exons within the gene.

One of the posttranslational modifying enzymes necessary for the production of collagen is protein disulfide isomerase, which, when combined with the alpha subunit of prolyl hydroxylase, forms a tetrameric protein isolated as prolyl hydroxylase. The gene for protein disulfide isomerase has been obtained from a human genomic library produced in a cosmid vector pcos 2EMBL 5 (Poustka et al., Proc Natl Acad Sci USA (1984) 81:4129-4133). The library was screened with cDNA fragments specific for human protein disulfide isomerase and several clones were obtained, at least two of which contained the entire gene (Tasanen et al., J Biol Chem 10 (1988) 263:16218-16224).

For use in the expression systems of the invention, this gene can be cut from the cosmid DNA with restriction endonucleases and ligated to a milk-specific protein promoter using a strategy similar to that used for the 15 construct of the heptoglobin-collagenase DNA.

In the event that the mammary cells are unable to provide suitable enzymes for posttranslational modification of the procollagen produced, the transgenic animals would need to be modified with expression systems 20 for these enzymes. Construction of these expression systems is analogous to that described herein for procollagen gene expression. The expression systems for the posttranslational enzymes are provided to the transgenic animal along with the expression systems for 25 the desired collagen product.

The choice of a promoter for expression in milk would preferably be from one of the milk-specific proteins, such as alpha S1-casein 5' and 3' regulatory sequences, which were fused to the human lactoferrin 30 cDNA, providing a construct that used the alpha S1-casein 35

promoter and signal sequence for the human lactoferrin gene. Another construct used to express a foreign protein in sheep milk consisted of the sheep beta-lactoglobulin promoter fused to human and antitrypsin gene 5 fragments (Wright et al., Biotechnology (1991) 9:830-833). A third promoter that has been used is the whey acid promoter, which was fused to cDNA for a modified version of human tissue plasminogen activator (Ebert et al., Biotechnology (1991) 9:835-838) and used to prepare 10 transgenic goats in whose milk human tissue plasminogen activator was expressed. The sequence of the gene is scanned for available unique restriction endonuclease sites, which are selected so that the functional gene containing the precise translation start site is 15 preserved in the mRNA.

In the event that it is desirable to provide posttranslational enzymes in the mammary cells, it is believed that the most important candidates are prolyl hydroxylase and protein disulfide isomerase. The gene 20 for the chick alpha subunit of prolyl hydroxylase has not yet been completely isolated, but is known to be as large as 50kb (R.A. Berg unpublished information). It is expected that the entire gene may be obtained from a human genomic cosmid library, as was done for the Col1A1 25 gene and the gene for protein disulfide isomerase. The cDNA for chick alpha subunit (Bassuk et al., Proc Natl Acad Sci USA (1989) 86:7382-7386) and human alpha subunit (Helaakoski, T., Proc Natl Acad Sci USA (1989) 86:4392-4396) have been described. Since the gene is not yet 30 available, the cDNA for the human alpha subunit for prolyl hydroxylase can be fused to the promoter for a milk-specific protein to produce a DNA construct for introduction into a transgenic animal.

Using these systems, animals are obtained which 35 secrete human collagen or procollagen into milk. The

gene encoding the desired procollagen chain is coupled to suitable control sequences which function in the mammary cells of mammalian species such as the regulatory sequences associated with the α S1 casein gene,
5 β -lactalbumin or α -lactalbumin genes, β -lactoglobulin or lactoferrin genes. Both 5' and 3' regulatory sequences can be used. The genes encoding the required posttranslational enzymes are similarly constructed into expression systems using mammary cell-specific regulatory sequences.
10

The resulting expression systems are microinjected using, for example, the technique described in U.S. Patent 4,873,191. The expression system constructs are amplified by PCR or cloning and purified by agarose gel electrophoresis. After electroelution, the concentration is adjusted to 1-10 μ g/ml and microinjected into the oocytes which are obtained from ovaries freshly removed from cows or other animals. The oocytes are aspirated from the follicles and allowed to settle before
15 fertilization with thawed frozen sperm capacitated with heparin and prefractionated by Percoll gradient to isolate the motile fraction.
20

The fertilized oocytes are centrifuged, for example, for eight minutes at 15,000 x g to visualize the pronuclei for injection and then cultured from the zygote to morula or blastocyst stage in oviduct tissue-conditioned medium. This medium is prepared by using luminal tissues scraped from oviducts and diluted in culture medium. The zygotes must be placed in the culture medium within two hours following microinjection.
25
30

35 Estrous is then synchronized in the intended recipient mammals such as cattle by administering coprostanol. Estrous is produced within two days and the embryos are transferred to the recipients 5-7 days after estrous.

Successful transfer can be evaluated in the offspring by Southern blot. By utilizing this system to effect the expression of the Col1A1 gene, for example, the offspring can be evaluated for the presence of the 5 Col1A1 gene by Southern hybridization using a Col1A1 gene derived probe.

Alternatively, the desired constructs can be introduced into embryonic stem cells (ES cells) and the 10 cells cultured to ensure modification by the transgene. The modified cells are then injected into the blastula embryonic stage and the blastulas replaced into pseudopregnant hosts. The resulting offspring are 15 chimeric with respect to the ES and host cells, and nonchimeric strains which exclusively comprise the ES progeny can be obtained using conventional cross-breeding. This technique is described, for example, in PCT Application WO91/10741, published 25 July 1991.

For production of the desired procollagen or 20 collagen in milk, expression systems for both the procollagen gene and the posttranslational enzyme-encoding genes must be present in the transgenic animal. There are several ways to achieve this.

First, the mammalian host may already produce the 25 required levels of posttranslational enzymes in the epithelial cells of the mammary glands. Alternatively, the constructs to be microinjected into eggs or transfected into ES cells may include a cocktail of the desired procollagen gene expression system along with the expression systems similarly constructed for, for 30 example, the prolyl hydroxylase and protein disulfide isomerase. The successful production of collagen in the milk can then be determined using antiprocollagen antibodies or by analysis of the milk for levels of hydroxyproline, a unique amino acid found in collagen as 35 a result of the activity of prolyl hydroxylase.

5 In another alternative, the expression systems for the procollagen gene and the expression systems for any needed posttranslational enzyme-encoding genes may be injected into different batches of fertilized eggs or
10 transfected into different batches of ES cells and used separately as described above to develop transgenic animals capable of expressing the procollagen or collagen genes and the posttranslational enzyme-encoding genes, respectively. These transgenic animals can then be
15 crossbred and the offspring evaluated for the ability to express both such systems. At least some of the offspring of such transgenic animals will be capable of producing both the collagen product and the posttranslational enzyme product.

15 In still another approach, fertilized eggs or ES cells may be prepared from transgenic animals already modified to have the capacity to express one or the other of the procollagen genes or the posttranslational enzyme-encoding genes. These eggs can then be microinjected or
20 the ES cells transfected with the expression system for the proteins lacking in the transgenic animal to develop into a transgenic animal containing expression systems for all of the required components.

25 Similarly, transgenic animals already modified with respect to one desired gene may be used as sources for the blastulas into which modified ES cells are implanted. Again, chimeric animals will result which can be used in cross-breeding to obtain offspring having genes for all of the desired proteins.

30 It may be noted that the expression systems for both of the particular posttranslational enzymes described above, if needed, must be provided essentially simultaneously since the enzymes function together as a tetrameric protein; as described above, the two α

subunits of prolyl hydroxylase must be associated with protein disulfide isomerase in order to be active.

When suitable transgenic mammals have been obtained by any of the foregoing methods, the procollagen or 5 collagen is secreted into the milk. The procollagen or collagen product of the transgenic mammal will be determined by the nature of the procollagen gene in the expression system provided. For homotriplexes, only a single gene is inserted. For production of 10 heterotriplexes, such as typical human collagen type I, either both the CollA1 and CollA2 genes are utilized in the original microinjection, or mammals transgenic for human CollA1 are crossbred with mammals transgenic for human CollA2. The type III collagen gene Col3A1 can be 15 used to prepare a transgenic animal and may be simpler because only one collagen polypeptide chain is required.

For the procollagen genes provided in the expression systems, procollagen is secreted into the milk if the required proteases for conversion to collagen are absent. 20 To the extent that these protease enzymes are absent from the secreting epithelial cells and are not provided for by recombinant systems, procollagen is secreted into the milk and can be recovered in a manner analogous to procedures that would be used for collagen *per se*. The 25 procollagen can also be converted before or after purification using specific proteases to cleave the prosequences as is known in the art. On the other hand, if the proteases are natively present intracellularly or are provided by recombinant systems, collagen will be 30 secreted directly. Depending on the levels of these enzymes, mixtures of procollagen and collagen may be obtained in the milk which can, if desired, be converted by treatment of the milk with proteases to convert all of the relevant molecules to collagen *per se*.

As described above, previous preparations of human collagen of a given type are always contaminated by the presence of alternative type collagens in view of the similarity of these materials and in view of the capacity 5 of native or other recombinant cells previously used to produce collagens encoded by their own genomes. By use of the method of the invention, it is possible to obtain collagen or procollagen of a given type free from coexpressed collagens or procollagens of alternative 10 types.

Purification of collagen or procollagen from milk is accomplished using their characteristic solubility and chemical properties. For example, milk may be acidified, causing milk-specific proteins such as casein to 15 precipitate and collagen or procollagen to remain in solution. The collagen or procollagen may be precipitated from acid solutions by the addition of salt, alcohol, or propylene glycol. (Miller, E.J. and Rhodes, R.K., Methods in Enzymology (1982) 82:33-64); Sage, H. 20 and Bernstein, P., ibid., 96-127.)

Claims

1. A method to prepare human collagen or human procollagen which method comprises:

5 recovering milk from the mammary glands of a nonhuman mammal which mammal has been modified to contain an expression system which comprises a coding nucleotide sequence encoding at least one human procollagen operably linked to control nucleotide sequences that effect expression specifically in milk protein-secreting
10 epithelial cells of said mammary glands under conditions wherein said coding nucleotide sequence is expressed to secrete human procollagen or human collagen into the milk of said mammal; and

15 recovering the human procollagen or human collagen from the milk.

2. An expression system for production of human procollagen or human collagen in milk which expression system comprises a coding nucleotide sequence encoding human procollagen operably linked to a promoter capable of specifically effecting expression in milk protein-secreting cells of mammary glands.

3. The expression system of claim 2 wherein the human procollagen is the pro- $\alpha 1$ chain of type I collagen or is the pro- $\alpha 1$ chain of type III collagen.

25 4. A fertilized nonhuman egg containing the expression system of claim 2.

5. A nonhuman embryonic stem cell modified to contain the expression system of claim 2.

6. A transgenic nonhuman mammal which comprises the expression system of claim 2.

7. The nonhuman mammal of claim 6 which has been further modified to contain at least one expression 5 system which effects the production of posttranslational modification enzymes for procollagen.

8. Milk containing human collagen or human procollagen.

9. A human procollagen or human collagen 10 composition containing only procollagen or collagen molecules of a single collagen type.

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US94/00740

A. CLASSIFICATION OF SUBJECT MATTER

IPC(5) :Please See Extra Sheet.

US CL :424/535; 435/69.1, 172.1, 172.3; 536/23.1, 23.5; 800/2

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 424/535; 435/69.1, 172.1, 172.3; 536/23.1, 23.5; 800/2

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

Please See Extra Sheet.

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	US, A, 4,873,316 (MEADE ET AL.) 10 OCTOBER 1989, see entire document.	1-9
Y	PROTEIN EXPRESSION AND PURIFICATION, Volume 1, issued 1990, L. Hennighausen, "The mammary gland as a bioreactor: Production of Foreign Proteins in Milk", pages 3-8, see entire article.	1-9
Y	PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCE, USA, Volume 85, issued August 1988, C.W. Pittius et al., "A milk protein gene promoter directs the expression of human tissue plasminogen activator cDNA to the mammary gland in transgenic mice", pages 5874-5878, see entire document.	1-9

Further documents are listed in the continuation of Box C. See patent family annex.

•	Special categories of cited documents:	
A	document defining the general state of the art which is not considered to be part of particular relevance	*T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
E	earlier document published on or after the international filing date	*X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
L	document which may throw doubt on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	*Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
O	document referring to an oral disclosure, use, exhibition or other means	*&* document member of the same patent family
P	document published prior to the international filing date but later than the priority date claimed	

Date of the actual completion of the international search

09 March 1994

Date of mailing of the international search report

31 MAR 1994

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INTERNATIONAL SEARCH REPORT

International application No.

PCT/US94/00740

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCE, USA, Volume 87, issued July 1990, A.L. Archibald et al., "High-level expression of biologically active human alpha-1-antitrypsin in the milk of transgenic mice", pages 5178-5182, see entire document.	1-9
Y	TRENDS IN BIOTECHNOLOGY, Volume 5, issued January 1987, A.J. Clark et al., "Pharmaceuticals from transgenic livestock", pages 20-24, see entire article.	1-9
P,Y	US, A, 5,227,301 (TURNER ET AL.) 13 JULY 1993, see entire document.	1-9
P,Y	US, A, 5,215,904 (GOULD ET AL.) 01 JUNE 1993, see entire document.	1-9
Y	US, A, 4,798,800 (TIMPL ET AL.) 17 JANUARY 1989, see entire document.	1-9
Y	METHODS IN ENZYMOLOGY, Volume 82, issued 1982, E.J. Miller et al., "Preparation and Characterization of the Different Types of Collagen", pages 33-69, see entire document.	1-9
Y	BIOTECHNOLOGY, Volume 9, issued September 1991, P.Krimpenfort et al., "Generation of Transgenic Dairy Cattle Using 'in vitro' embryo production", pages 844-847, see entire document.	1-9
Y	THE JOURNAL OF BIOLOGICAL CHEMISTRY, Volume 263, number 26, issued 15 September 1988, S-T. Lee et al., "Construction of a Full-Length cDNA Encoding Human Pro-alpha2(I) Collagen and its Expression in Pro-alpha2(I)-deficient W8 rat cells", pages 13414-13418, see entire document.	1-9
Y	THE JOURNAL OF BIOLOGICAL CHEMISTRY, Volume 267, number 7, issued 05 March 1992, A. Torre-Blanco et al., "Copolymerization of Normal Type I Collagen with Three Mutated Type I Collagens Containing Substitutions of Cysteine at Different Glycine Positions in the alpha1(I) chain", pages 4968-4973, see entire document.	1-9
Y	CELL, Volume 71, issued 11 December 1992, J.D'Armiento et al., "Collagenase Expression in the Lungs of Transgenic Mice Causes Pulmonary Emphysema", pages 955-961, see entire document.	1-9

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US94/00740

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCE, USA, Volume 82, issued May 1985, K.S.E. Cheah et al., "Identification and characterization of the human type II collagen gene (COL2A1)", pages 2555-2559, see entire article.	1-9
Y	THE JOURNAL OF BIOLOGICAL CHEMISTRY, Volume 266, number 22, issued 05 August 1991, L. Ala-Kokko et al., "Expression of a human cartilage procollagen gene (COL2A1) in Mouse 3T3 cells", pages 14175-14178, see entire document.	1-9
Y	PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCE, USA, Volume 88, issued September 1991, P. Vandenberg et al., "Expression of a partially deleted gene of human type II procollagen (COL2A1) in transgenic mice produces a chondrodysplasia", pages 7640-7644, see entire document.	1-9
Y	THE JOURNAL OF BIOLOGICAL CHEMISTRY, Volume 263, number 31, issued 05 November 1988, K. Tasanen et al., "Characterization of the human gene for a polypeptide that acts both as the beta subunit of prolyl 4 hydroxylase and as protein disulfide isomerase", pages 16218-16224, see entire document.	1-9
Y	THE JOURNAL OF BIOLOGICAL CHEMISTRY, Volume 264, number 34, issued 05 December 1989, D.S. Greenspan et al., "High levels of Expression of Full length human Pro-alpha2(V) collagen cDNA in Pro-alpha2(V)-deficient Hamster Cells", pages 20683-20687, see entire document.	1-9
Y	THE JOURNAL OF BIOLOGICAL CHEMISTRY, Volume 266, number 2, issued 15 January 1991, A.S. Olsen et al., "High levels of expression of a minigene version of the human Pro alpha1(I) Collagen gene in stably transfected mouse fibroblasts", pages 1117-1121, see entire document.	1-9
Y	THE JOURNAL OF BIOLOGICAL CHEMISTRY, Volume 267, number 16, issued 05 June 1992, K. Tasanen et al., "Promoter of the gene for the multifunctional Protein Disulfide Isomerase Polypeptide", pages 11513-11519, see entire article.	1-9
Y	SCIENCE, Volume 246, issued 08 December 1989, D. Hanahan, "Transgenic Mice as Probes into Complex Systems", pages 1265-1275, see entire article.	1-9
Y	THE NEW ENGLAND JOURNAL OF MEDICINE, Volume 311, number 6, issued 09 August 1984, D.J. Prockop et al., "Heritable Diseases of Collagen", pages 376-386, see entire article.	1-9

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US94/00740

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	TRENDS IN GENETICS (TIG), Volume 2, issued January 1986, E.J. Robertson, "Pluripotential stem cell lines as a route into the mouse germ line", pages 9-13, see entire article.	1-9

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US94/00740

A. CLASSIFICATION OF SUBJECT MATTER:
IPC (5):

A23C 9/00; A23J 1/00; C07H 15/00; C07K 1/00, 3/00, 13/00, 15/00, 17/00; C12N 5/00, 15/00; C12P 21/06

B. FIELDS SEARCHED

Electronic data bases consulted (Name of data base and where practicable terms used):

Electronic Databases: APS, MEDLINE, CA

Search terms: collagen?, procollagen?, mammary, mammary, gland?, transgen?, prolylhydroxylase, prolyl (2a) hydroxylasse, bovine, cow, ovine, sheep, berg, richard a/au, berg ra/au, therap?, human, administ, recomb?, promoter?